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Functional Differentiation of Type II and Type I Collagen Articular Models in Synovial Fluid Film Formation and Recombinant Equine Lubricin Retention

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ABSTRACT

Collagen type II (Col-II) and collagen type I (Col-I) are major components of articular cartilage present at different ratios at its surface. Understanding how each of these components mediates the assembly of molecular films derived from synovial fluid (SF), the lubricant of synovial joints, is critical to explain the loss of mechanical performance in pathological conditions, guide the design of biomaterial implants meant to be in contact with SF, and develop molecular therapies to restore SF properties. This work demonstrates that Col-II articular surface model assists in scaffolding SF-derived films, while Col-I model lacks SF film scaffolding capabilities. However, when Col-II and Col-I are exposed to recombinant equine lubricin (rEqLub), an analog of the major boundary lubricant in SF, both adsorbed and retained similar amounts. These insights, deduced from quartz crystal microbalance with dissipation, diffuse reflectance circular dichroism, and atomic force microscopy, reveal possible mechanisms underlying the loss of mechanical performance of synovial joints in pathology, where Col-I becomes the major collagenous component of the articular cartilage surface, as well as considerations for designing functional biomaterial implants. Furthermore, this work reinforces the idea of rEqLub as an intra-articular osteoarthritis therapy with the ability to bind to Col-II and Col-I, irrespectively.

1 | Introduction

Synovial joints are defined by their mobility, where bone surfaces are covered by articular cartilage that enables low-friction movement and repetitive mechanical loading, facilitated by synovial fluid (SF) lubrication. At the interface between the surface of the articular cartilage extracellular matrix (AC-ECM) and SF, a

thin film of SF components existing as a gel-like layer dissipates loads and reduces wear, damage, and friction forces in boundary lubrication mode [1, 2]. The degree of hydration at this interface has been proposed to be, in part, responsible for the tribological robustness of synovial joints [3–5], along with synergistic interactions between glycosaminoglycans (GAGs, e.g., hyaluronic acid, HA), phospholipids (PLs, e.g., phosphatidylcholine, PC),

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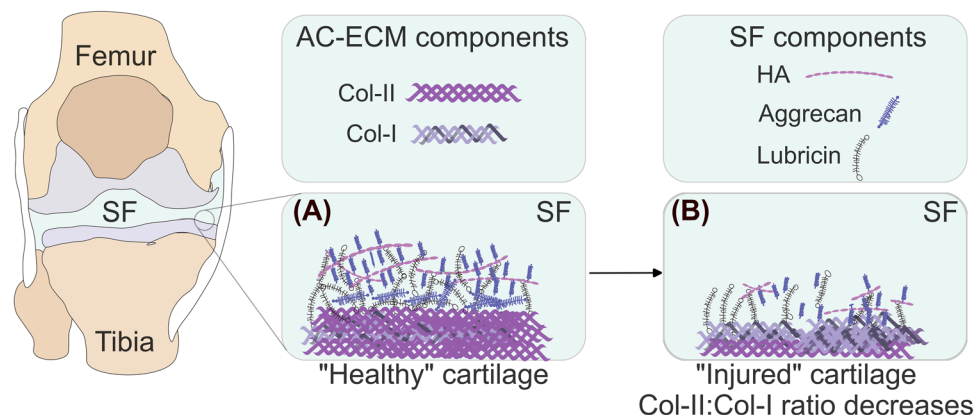


FIGURE 1 | Schematic of a knee (prototypical synovial joint) and the regions in which this study focuses, i.e., the interface between SF and AC-ECM. (A,B) represent “healthy” and “injured” cartilages, respectively, considering only the collagenous components of the AC-ECM and some SF components.

and glycoproteins (GP, e.g., lubricin, Lub), whose individual roles (or low-order mixtures) have been widely investigated [4, 6–20]. Another key mechanism proposed for boundary lubrication involves steric-entropic repulsive forces that result from the flexibility of coiled molecules resisting conformational restriction upon confinement, as has been demonstrated for Lub alone and Lub mixed with HA [18, 21]. Thus, recombinant equine Lub (rEqLub) analog has been proposed as a key molecule with potential to treat synovial joint diseases, such as osteoarthritis [22–25].

While these studies have shaped our understanding of synovial joint nanomechanics, fully capturing SF synergies on a molecular level is essential to enable the efficient design of the next generation of intra-articular osteoarthritis therapies. Collagen type II (Col-II) is the main component of healthy AC-ECM (Figure 1A), while collagen type I (Col-I) is also present at the surface of articular cartilage, with its content increasing in damaged cartilage or fibrocartilaginous repaired tissue (Figure 1B) [26–28]. Some studies have shown that SF components, such as Lub, can bind to Col-II at the molecular level, forming anti-adhesive and wear-protecting layers [29, 30]. Also, solid-phase binding assays have shown that rEqLub can bind to Col-II and fibronectin (FN), another molecular component of the AC-ECM [31].

Efforts have been made to fabricate matrices of Col-II [32], Col-I [33], and mixtures of both [34] in order to support cartilage repair [35], screening of preclinical drugs [36], and understanding how both of these collagens interact with key SF components at the molecular level [29, 30, 37]. However, the roles of Col-II and Col-I in the supramolecular assembly of a film derived from the totality of SF components, as well as the resulting nanomechanical properties of that film, such as normal (load-carrying) and lateral (friction) compliance, remain unexplored. Furthermore, it is well established that the constituents of extracellular matrices appear in a range of molecular conformation distributions, which are affected and altered by disease and mechanical stimuli [38–45]. How AC-ECM alterations, such as molecular conformations of collagens (e.g., II or I), and surface morphologies, may affect the adsorption of SF components, and thus SF film assembly and mechanics, is yet another open question. This is important to guide effective pathophysiological treatments, including the

design and application of biomaterials for cartilage repair and synthetic SFs, based on, for example, rEqLub.

Col-II and Col-I are fibril-forming collagens and consist of three α -polypeptide chains that form a right-handed triple helix, generating a collagen molecule [46]. While mostly similar, they differ in key molecular features that give each its own characteristics. On one hand, both collagen molecules have a molecular weight of ~ 280 – 300 kDa, a contour length of ~ 300 nm, and a diameter of ~ 1.5 nm [46, 47]. However, Col-II is a homotrimer consisting of three $\alpha_1(\text{II})$ polypeptide chains [46], whereas Col-I is a heterotrimer consisting of two $\alpha_1(\text{I})$ and one $\alpha_2(\text{I})$ polypeptide chains, encoded by different genes. $\alpha_1(\text{I})$, $\alpha_1(\text{II})$, and $\alpha_2(\text{I})$ have different amino acid sequences, more specifically, different interruptions of the typical Gly-X-Y amino acid sequence [48]. These chains assemble onto triple helices to create collagen molecules, which then assemble with a staggered arrangement into fibrils whose length and diameter in vitro are in the order of microns and nanometers, respectively [49–51]. In the AC-ECM, one of the principal roles of collagen molecules is to provide mechanical integrity.

In solution, the stability of collagen triple helices, and thus their tertiary structure, is strongly dependent on the environmental conditions, such as pH [50, 52, 53]. In this work, molecular films of Col-II and Col-I were formed by controlling their bulk conformations via solution pH adjustment, to either pH ~ 7 or pH ~ 4 , followed by chemical immobilization onto NH_2 -terminated self-assembled monolayers (SAMs) on gold-coated quartz crystals via glutaraldehyde activation [29, 54]. This process yielded immobilized collagen films with different nano-morphologies and nano-mechanics as evidenced by measurements of hydrated film mass density, Young's modulus, film elastic compliance, film morphology, and film molecular conformation. To characterize these films, quartz crystal microbalance with dissipation (QCM-D), atomic force microscopy (AFM) in a liquid environment, and diffuse reflectance circular dichroism (DRCD) were used. Furthermore, the efficacy of these films to retain SF and rEqLub was assessed via QCM-D.

Results revealed differences in the assembly of SF-derived films on Col-II compared to Col-I surfaces. Col-II retained significantly

larger amounts of SF components than Col-I. On the other hand, all Col-II and Col-I films retained similar amounts of rEqLub, suggesting that the binding affinity of rEqLub is similar for Col-II and Col-I under the studied conditions. These results give insight into how altered AC-ECM loses SF supramolecular assembly capabilities when Col-II is replaced by Col-I, with possible implications in loss of load-bearing potential [13] and consequently, loss of mechanical performance. On the other hand, our findings suggest that the tested rEqLub could be explored for therapeutic use in injured cartilage or for supplementation of lubrication-deficient SF.

2 | Results and Discussion

2.1 | Saturation of Crystals was Reached with 50 $\mu\text{g/mL}$ Col-II and Col-I Solutions

First, to determine the bulk concentration of collagen (II and I) that saturates the SAM-functionalized gold crystals, chemisorption experiments were performed with QCM-D at collagen concentrations ranging from 10 to 100 $\mu\text{g/mL}$. Col-II saturated crystals above 50 $\mu\text{g/mL}$, whereas Col-I saturated crystals at all tested concentrations (Figure S1A). Shear compliance did not show dependence over the tested range of concentrations for any of the collagens (Figure S1B). Based on this, 50 $\mu\text{g/mL}$ was chosen as the concentration for collagen film formation.

2.2 | Collagen at Neutral pH Exists in a Partially Denaturated State

It is well established that environmental conditions, such as pH, have an impact on collagen molecular conformation, modulating the stability of the collagen triple helix [50, 52, 55, 56]. To assess the conformational state of Col-II and Col-I prior to surface immobilization, CD spectra in PBS solution at pH 7 and pH 4 were acquired. Col-II presented a negative peak at ~ 197 nm, with a zero-rotation point at ~ 213 nm, and no positive maximum at ~ 222 nm (as would have been expected for Col-II with a perfect triple helical structure [57]). The negative peak at 197 nm indicates the presence of α -helices, likely originating from the individual polypeptide chains of Col-II (Figure 2A). In contrast, Col-I at pH 7 and pH 4 showed clear triple-helix signatures in the CD spectrogram (Figure 2A), with a negative peak at 197 nm, a zero-rotation point at 213 nm, and a positive maximum at 222 nm, consistent with previous reports [56, 58]. However, pH 7 induced conformational changes relative to pH 4, as evidenced by a decrease in the MRE amplitude of both peaks at pH 7. In the case of Col-II, R_{pn} is lower than that of Col-I in both pH 4 and pH 7, suggesting partial denaturing of the triple helical structure, regardless of pH. While the R_{pn} of Col-II at pH 4 and 7 are comparable, it should be noted that the absolute values of the positive and negative peaks decreased at pH 7, suggesting a certain degree of overall disordering taking place in the switch from acidic to neutral pH. While the R_{pn} as a ratio of absolute peak values, is in principle independent of sample concentration, a uniform flattening of the curve would not be reflected in the value, even though it is a strong indicator of collagen degradation [59]. In view of this, Col-I at pH 4 appears to exhibit the canonical triple-helical structure, while Col-II is found

to exist in a partially degraded state. Furthermore, pH 7 seems to trigger partial degradation of both Col-I and Col-II.

2.3 | Col-II Film Assembly was pH-Dependent

As demonstrated by CD spectra (Figure 2A), pH affected the molecular conformation of Col-II and Col-I in bulk. To test whether pH-induced conformational changes affected the mass density and mechanical properties of collagen films, QCM-D was used. Collagen nanofilms were characterized by estimating their hydrated mass densities, as determined by their mass density, m_f , (Figure 2B), and shear-dependent film compliances, J'_f (Figure 2C).

Figure 2B shows the m_f of the pre- and post-wash of immobilized Col-II and Col-I films at pH 7 and 4, taken from at least $N = 6$ independent measurements. Inferring from two-way ANOVA tests (Table S2), collagen type and film formation pH were both variables that significantly impacted m_f . No differences were observed between pre- and post-wash collagen films, indicating that the molecular content initially present in the films was retained after the PBS washing steps. Thus, the discussion for m_f focuses on the post-wash conditions. Additional t -tests with Holm-Bonferroni correction revealed that Col-II films at pH 7 formed denser (thicker) films than those at pH 4, but this was not the case for Col-I. The mean film thicknesses of Col-II films at pH 7 were $\sim$ three times higher than those of films formed at pH 4. Lastly, the difference in film thickness between Col-II and Col-I formed at pH 7 was not statistically significant. This is not the case for film thicknesses at pH 4, where Col-II films were $\sim$ four times thinner than Col-I.

It is evident that pH impacted the m_f of Col-II films more than m_f of Col-I films. This might arise from several factors, including molecular differences (homotrimer in Col-II vs heterotrimer in Col-I), differences in the interruption of the Gly-X-Y domains, higher-order structure, and differences in isoelectric points (I_p). Both Col-II and Col-I have an overall positive charge in acidic conditions (pH 4), as their I_p values are ~ 8 and ~ 7 , respectively, resulting in a larger difference in surface charge for Col-II as compared to Col-I between the explored pH values. These factors likely led to differences in packing densities and, thus, hydration states between Col-II and Col-I films.

For films deposited at pH 4, a subtle swelling was observed when transitioning to pH 7 (pH 4 to 7 conditions in Figure 2B), but it was not enough to be considered statistically relevant based on t -test analysis.

2.4 | Col-II Formed Less Compliant Films at pH 4 and pH 7 than Col-I when Deposited at pH 7

To further understand the mechanical properties and dissipative nature of the collagen films, we computed their shear-dependent thin film compliance, J'_f . For all conditions, no significant relative changes between pre- and post-wash for J'_f were measured. However, J'_f of Col-II films formed at pH 7 were statistically less compliant than Col-I films formed at pH 7, and Col-II films formed at pH 4 were less compliant than Col-I films formed at

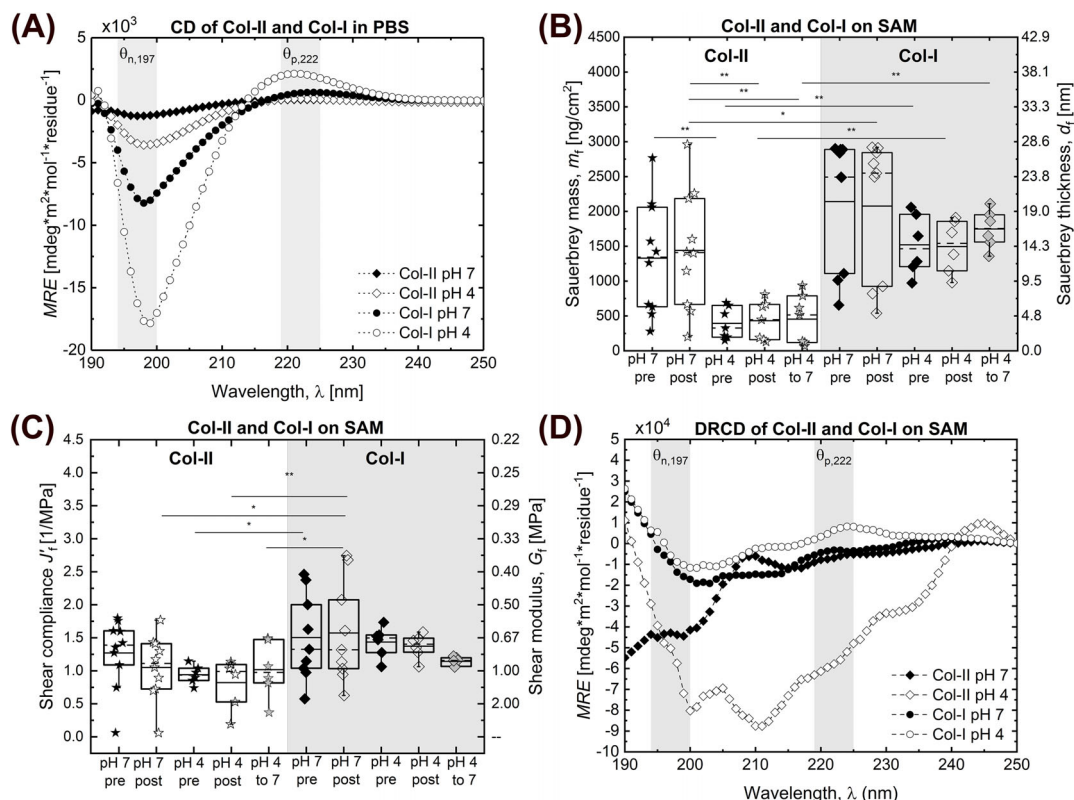


FIGURE 2 | (A) Far UV CD spectra of Col-II and Col-I at pH 7 and 4 in bulk. (B) mass density and (C) thin film compliance of Col-II and Col-I immobilized on SAM before (pre) and after (post) wash with PBS at their respective pH, followed by a PBS pH 7 wash for the samples deposited at pH 4 (shown as pH 4 to 7), quantified via QCM-D. For conditions starting at pH 4, an additional wash with PBS at pH 7 was performed. Box-and-whisker plots in (B,C) display the mean (solid line), median (broken line), lower and upper quartiles (box), and minimum/maximum values excluding outliers with a coefficient of 1.5 (whiskers). Each data point represents an independent replicate. (D) Far UV DRCD spectra of Col-II and Col-I deposited from PBS at pH 7 and 4, immobilized on SAM.

pH 7. After film formation at pH 4, changing the pH to 7 did not result in J_f' changes.

The means and standard error of the means (SEMs) for the QCM-D measurements are found in Table S1. Statistical analysis results, based on two-way ANOVA and pairwise comparisons with Holm-Bonferroni corrections, for m_f and J_f' are shown in Tables S2 and S3 and the Supporting Information “QCM-D_Pairwise_Comparisons.xlsx”. The trends for the QCM-D measured properties are similar for the case of physisorbed proteins on bare Au, although the values of adsorbed mass density are, in general, higher (Figure S2).

To obtain further information on the molecular conformation in the formed films, DRCD spectra of the chemisorbed films were collected. Most of the structural signatures were lost for both collagens when immobilized to the surfaces at acidic pH (Figure 2D). We hypothesize that this is due to the chemical immobilization process itself disrupting the collagen molecular structure. In our previous work on fibronectin films, we observed that the immobilization strategy significantly impacts the DRCD signal [54]. Nevertheless, DRCD spectra of Col-I at pH 4 and pH 7 agree with bulk CD data, as canonical Col-I peak amplitudes were more pronounced compared to Col-II. As in bulk, peak amplitudes were also less pronounced at pH 7 than at pH 4, again indicating some degradation of the Col-I structure at pH 7.

The above-mentioned photo- and acoustic-spectroscopy techniques provided conformational and nanorheological information on the impact of pH on Col-II and Col-I film formation. Based on these results, it is evident that the formation pH led to collagen films with different molecular conformations and properties, thereby accessing a wider range of collagen conformations than otherwise would have resulted from depositing at a single pH value, refining the articular cartilage surface model presented here. It is important to remember that molecular conformations in extracellular matrices appear in a distribution of conformations [41], most likely the case with articular cartilage surface as well, and thus the importance of capturing potentially different molecular conformations at the surface of articular cartilage. To further obtain morphological and nanomechanical information of the films, atomic force microscopy (AFM) in liquid was employed, as discussed next.

2.5 | Col-II and Col-I Formed Continuous Films with Similar Nanoscale Morphologies Across Conditions

To validate that the collagen precursor films were continuous and gain insights into their morphologies, the surfaces of nanofilms fully immersed in liquid (PBS at pH 7) were imaged using AFM in QI mode (Figure 3). Col-II and Col-I, regardless of the deposition

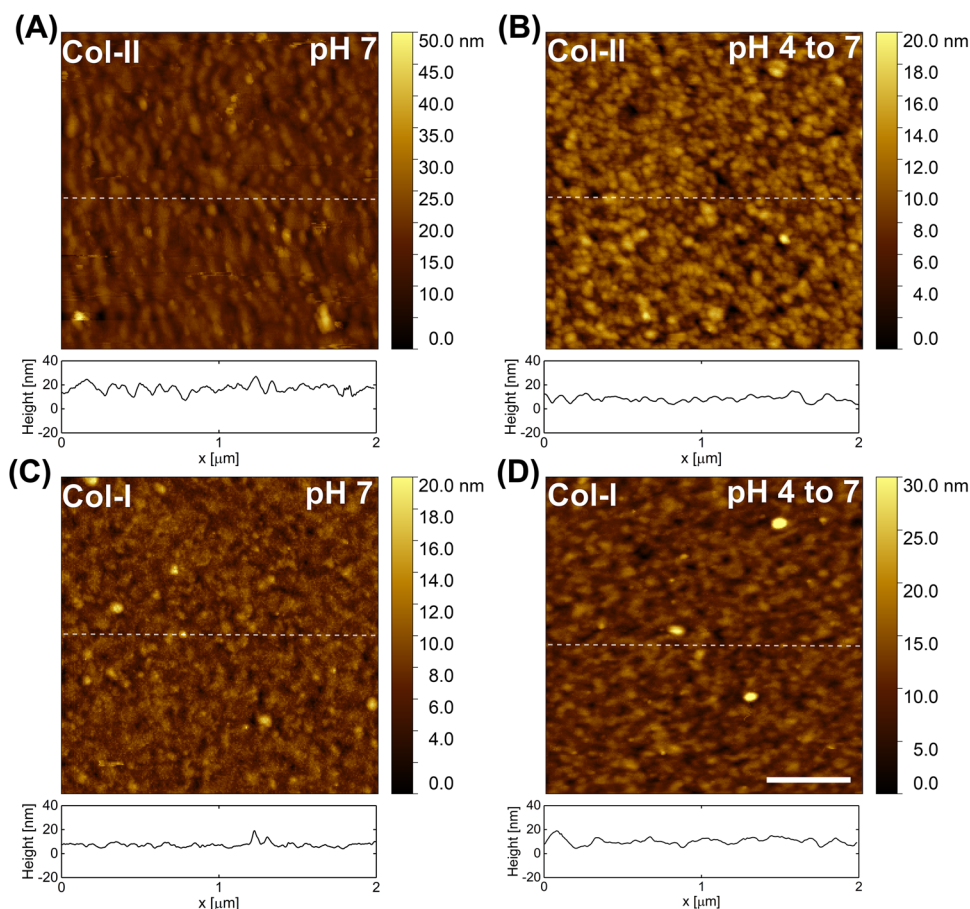


FIGURE 3 | Representative AFM height maps of (A) Col-II pH 7, (B) Col-II pH 4, (C) Col-I pH 7, (D) Col-I pH 4. All conditions were measured in a PBS liquid environment at pH 7. Representative cross sections of each map are shown below them. Scale bar in (D) applies to all micrographs of this figure and is 500 nm. At least two independent replicates were measured for each condition, showing similar results.

pH, formed continuous films with globular features, suggesting that the collagen is present in an amorphous or aggregated state and does not form fibrillar structures. These morphologies were expected and have been reported [30, 60]. Additionally, larger $5 \times 5 \mu\text{m}^2$ area scans are presented in Supporting Information (Figure S3).

$2 \times 2 \mu\text{m}^2$ micrographs of Col-II films deposited at pH 7 show surfaces with an RMS roughness of 4 nm, while those deposited from pH 4 had an RMS roughness of 3 nm. Col-II formed at pH 7 presented slightly more elongated structures that appeared to have a preferred orientation along the slow axis of the AFM scan. These elongated structures can also be seen in the $5 \times 5 \mu\text{m}^2$ area scans (Figure S3C). Col-I films deposited at pH 7 had an RMS roughness of 3 nm, while those deposited at pH 4 had an RMS roughness of 2 nm. Similar morphologies and RMS roughness values of physisorbed Col-II films on gold under similar experimental conditions have been reported in the literature [30].

In environments with relatively high ionic strength, it has been shown that salt ions screen intermolecular interactions between Col-II triple helices, negatively impacting collagen microfibril formation [53, 61]. This effect begins to appear at physiological ionic strengths (~ 150 – 200 mM), a range that includes the ionic strength of our PBS buffer (~ 160 mM). This can be a reason for

the lack of large microfibril structures in the AFM micrographs shown (in addition to the low concentrations of Col-II and Col-I of 0.05 mg/ml used in this study). To further understand the normal nanomechanics of Col-II and Col-I films, an independent set of samples was characterized by AFM force-volume mapping, as discussed next.

2.6 | Col-II Formed Stiffer Films than Col-I when Deposited at pH 4 but not at pH 7

Next, we tested if the trends and magnitudes observed by nanorheological properties measured via QCM-D (shear-compliance) held for nanoindentation via AFM (Figure 4).

Nanomechanical characterization based on force-volume mapping performed on each of the precursor collagen films revealed differences in Young's modulus (or compliance) between Col-II and Col-I films deposited from pH 4, but not between Col-II and Col-I films deposited at pH 7. Furthermore, the distribution of the Young's moduli for Col-II deposited at pH 4 showed a normal distribution, with a mean and standard deviation (STD) of 3.00 ± 1.93 MPa, while for Col-I deposited at pH 4, the distribution of Young's moduli resembles more a right-skewed normal distribution with a mean and STD of 1.39 ± 1.16 MPa. That is, Col-II films deposited at pH 4 showed a mean of E of $\sim$

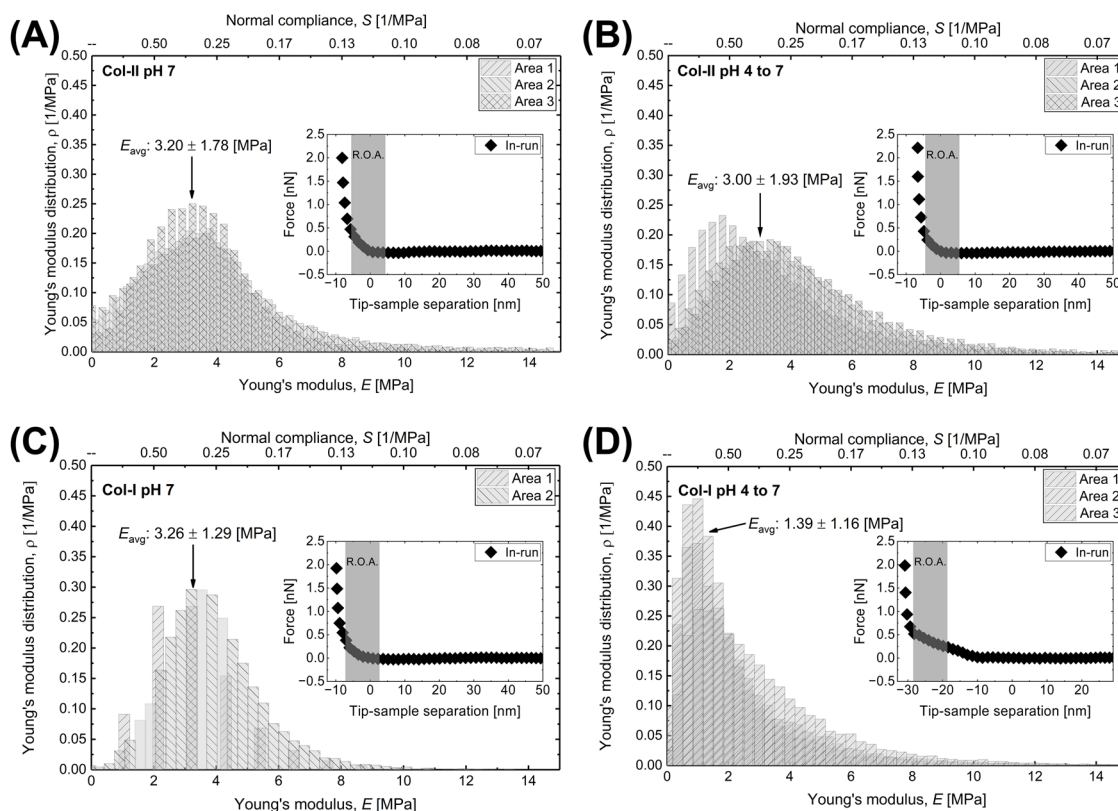


FIGURE 4 | AFM Young's moduli distributions of (A) Col-II pH 7, (B) Col-II deposited at pH 4 and measured at pH 7 (pH 4 to 7), (C) Col-I pH 7, (D) Col-I pH 4 to 7. All conditions were measured in a PBS liquid environment at pH 7. A secondary horizontal axis (top) reports normal compliance, S , as the inverse of the Young's moduli ($S=1/E$). Insets are representative force-distance curves for each condition, with their region of analysis (R.O.A.) used to extract E . E_{avg} indicates the average of the means obtained for each individual distribution, as shown in Figure S7. Each "Area" histogram represents an independent technical replicate.

double than that of E for Col-I. For films formed from pH 7, the difference is barely noticeable, having means and STDs of 3.20 ± 1.78 MPa and 3.26 ± 1.29 MPa for Col-II and Col-I, respectively. Notice that the mean and STD reported are the average of the mean and STD of the independent areas for each condition, which were all independently fitted with Gaussian functions (Figure S4). All collagen films, however, showed some degree of hysteresis between the in- and out-runs, as shown in the representative force-distance curves (Figure S5). The response of collagen films to the interaction with the AFM probe (which had a maximum ~ 2 nN, for a radius of curvature ~ 20 nm) was not purely elastic, as the recovery (if any) was not immediate. This is not surprising, as collagen is known to have a viscoelastic response under nanoindentation [62]. These findings, however, show that this characteristic of collagen is most likely present even when the triple-helical structure is largely lost (in agreement with DRCD). Young's moduli values of these nanofilms are consistent with previously reported values for collagen gels, obtained from nanoindentation [63].

Connecting the nano-rheological properties obtained from QCM-D and the nano-mechanics for AFM, if the collagen films are considered isotropic, homogeneous, and linearly elastic, the Young's modulus E should be approximately three times the shear modulus G_r , which is in good agreement with the elasticity measurements of Col-II. However, this is not the case for Col-

I, suggesting a deviation from an isotropic, homogeneous, and linearly elastic assumption (Table S4).

2.7 | Col-II Acted as a Scaffold for Dilute Synovial Fluid (dSF) and Recombinant Equine Lubricin (rEqLub), while Col-I Adsorbed and Retained rEqLub Only

To assess the capability of the Col-II and Col-I films to scaffold the supramolecular assembly of dSF and rEqLub films, QCM-D was used. Collagen films formed in the QCM-D system, as described above, were further exposed to dSF rEqLub. The m_f of the adsorbed and retained dSF or rEqLub on the collagen films was quantified (Figure 5).

Significant difference in dSF adsorption on Col-II deposited at pH 7 versus pH 4 was measured, for pre- and post-wash m_f values, suggesting that the Col-II molecular conformational differences embedded in the films (Figure 2B,D) do impact the supramolecular assembly of dSF films. Unlike Col-II films, Col-I at pH 7 showed m_f values close to zero or negative (within detectable limits of the QCM-D). These measurements are reported as "little or no adsorption" (Table S5). One plausible interpretation of the negative m_f values is that dSF molecules with higher affinities for Col-I and smaller molecular weight displaced

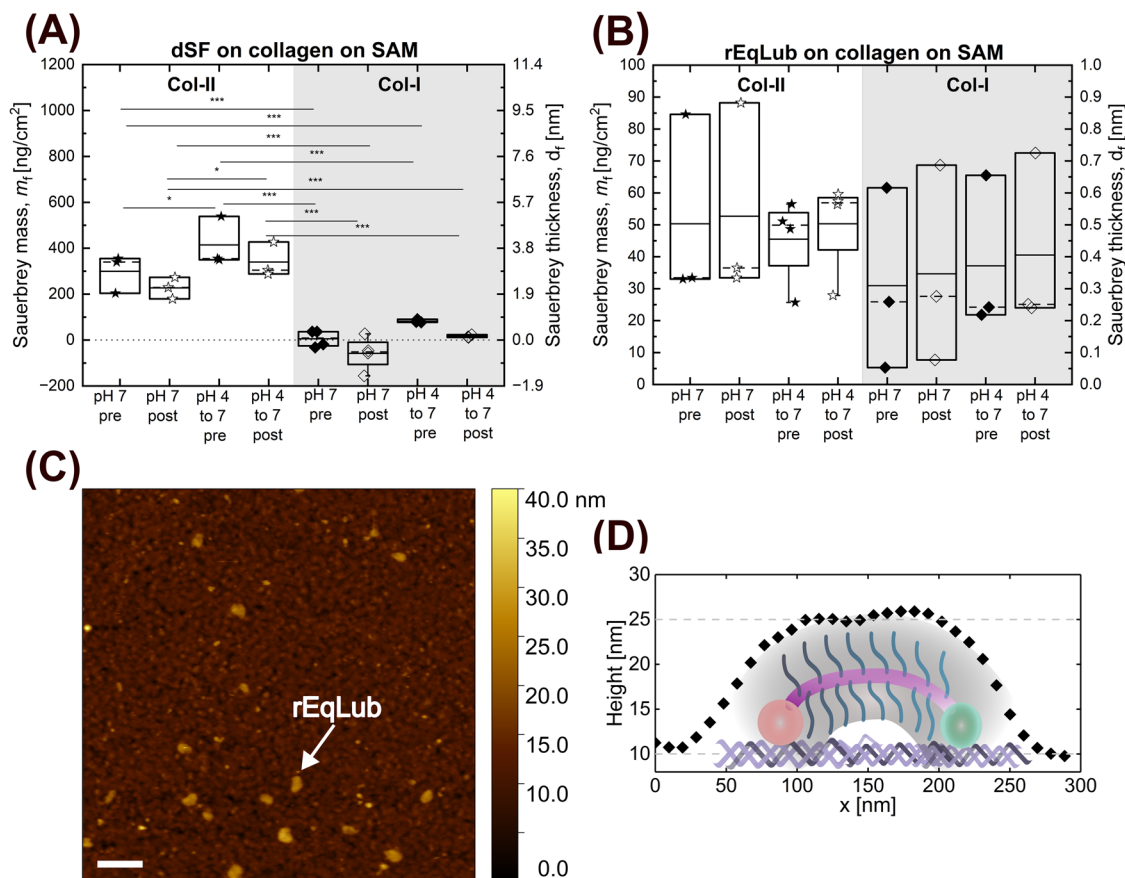


FIGURE 5 | QCM-D computed m_f of (A) dSF and (B) rEqLub adsorbed and retained layers on collagen precursor films. Note that the vertical axes are on different scales for (A,B). Box-and-whisker plots in (A) and (B) display the mean (solid line), median (broken line), lower and upper quartiles (box), and minimum/maximum values excluding outliers with a coefficient of 1.5 (whiskers). Each data point is an independent replicate. (C) AFM height image (contact mode) of rEqLub on a Col-I film. Scale bar is 500 nm. (D) Average height profile of 5 rEqLub molecules of (C), to visualize a possible conformation of rEqLub molecules attached to the surface.

adsorbed BSA (from the previous blocking step), resulting in a thinner Sauerbrey film than the initial one. This phenomenon is known as the Vroman effect [64]. One such family of molecules is phospholipids, which are abundant in SF and are proposed to play a key role in mediating cartilage surface properties [4, 65–67]. It is important to note that serum albumin (similar to the blocking molecule), is a major constituent of SF [11]. On Col-I films deposited at pH 4, unlike those at pH 7, the m_f of dSF was positive. After the wash, this amount significantly decreased, indicating a loss of most of the adsorbed mass. Both adsorbed and retained dSF films on Col-I at pH 4 to 7 were, however, significantly smaller than the m_f for dSF on either Col-II condition, and no statistically significant differences were observed.

Overall, the type of collagen mediates the scaffolding of dSF, as reflected by dSF m_f (or, analogously, d_f thickness of dSF films formed), supported by two-way ANOVA and pairwise comparisons, as shown in Table S6 and Supporting Information “QCM-D_Pairwise_Comparisons.xlsx”. This finding can contribute to understanding loss in the mechanical performance of compromised synovial joints, considering that the Col-II to Col-I ratio decreases in pathologies such as osteoarthritis and rheumatoid arthritis [48]. Furthermore, this finding can assist in

the design of biomaterials for cartilage repair or bioactive coatings for orthopedic implants that are directly exposed to SF.

Next, we explored the role of Col-II and Col-I in scaffolding rEqLub, an analog of lubricin (Lub), also known as superficial zone protein (SZP) or PRG4, a molecule identified as a key regulator of friction and synovial joint mechanics [13, 68] present in SF, and proposed as a therapy for pathological synovial joints [22]. A key requirement of boundary lubricants is the ability to adsorb and remain attached to the surface, as validated for rEqLub via QCM-D.

2.8 | Col-II and Col-I Adsorbed and Retained Similar Amounts of rEqLub Molecules for All Conditions

We quantified the adsorption of rEqLub directly on Col-II and Col-I films formed at pH 7 and 4 (Figure 5B). For all conditions tested, no statistically significant differences in their m_f were observed. The fact that rEqLub adsorbed to both collagens was expected, as previous studies have discussed the capability of purified and recombinant lubricin to adsorb to Col-II and Col-I precursor films [28, 30, 69]. This contrasts with dSF m_f findings,

suggesting that the assembly of dSF films on the precursor films was not driven solely by lubricin, but by intermolecular synergies among SF components, as has been reported for HA and Lub [18], HA and PLs [4], and HA with Aggrecans [70]. m_f values for rEqLub are summarized in Table S5.

Based on the conversion from m_f to d_f (Sauerbrey thickness) used in this study, rEqLub films across conditions should have thicknesses less than a nanometer if they were continuous and homogeneous. However, this is not realistic, as a single hydrated lubricin molecule is expected to have a cross-sectional diameter of ~ 10 nm. Our results suggest that rEqLub molecules did not form continuous, homogeneous films, but rather, rEqLub decorated collagen surfaces discretely. Indeed, AFM imaging of rEqLub on Col-I supports the interpretation of a discretely decorated collagen film (Figure 5C). This has been observed and reported for rLub (different from rEqLub one used in this study) on Col-II via AFM as well [30], with the same protein concentrations used in our study.

Furthermore, the average cross-section of five different rEqLub molecules on Col-I is presented in Figure 5D, along with a schematic of a possible conformation of the rEqLub monomers. This is, assuming that the rEqLub monomers are either lying flat on the Col-I surface, or forming loops, assuming they are attached through their C- and N- termini to the underlying collagens. Another possibility is that of rEqLub multimers interconnecting through their N- termini, remaining bound to the Col-I surface via their C- termini only, as has been proposed previously for lubricin binding to fibronectin [11]. The exact way in which lubricin decorates AC-ECM surfaces remains an open question. Results of two-way ANOVA of the adsorbed m_f comparisons for rEqLub are presented in Table S7, with pairwise comparisons shown in the Supporting Information “QCM-D_Pairwise_Comparisons.xlsx”.

2.9 | dSF Disrupted the Mechanical Integrity of Col-I Films

Lastly, to gain insights into the compliance (which incorporates contributions of the QCM-D dissipation channel to the modeling, while m_f does not) of the dSF and rEqLub on Col-II and Col-I, shear compliance J_f' of dSF and rEqLub on Col-II and Col-I, was quantified. Results of the post-wash conditions are shown in Figure 7. Pre- and post-wash J_f' values for dSF and rEqLub on Col-II and Col-I are presented in Table S8. Results of J_f' of dSF and rEqLub pairwise comparisons are presented in Supporting Information “QCM-D_Pairwise_Comparisons.xlsx”.

J_f' post-wash values for collagen films with dSF revealed statistically significant differences between Col-II pH 4 to 7 and Col-I pH 7 only (Figure 6). These differences, however, are most likely dominated by Col-II precursor film J_f' (Figure 2C). When comparing the films of collagen against collagen films with adlayers (dSF or rEqLub), a significant difference between J_f' of Col-I pH 4 to 7 and dSF + Col-I pH 4 to 7 was revealed, with a more compliant system when SF components were adsorbed to the precursor Col-I film. For Col-I pH 4 to 7 films with rEqLub, there was no statistically significant difference in any condition compared with their respective measurements with collagen alone.

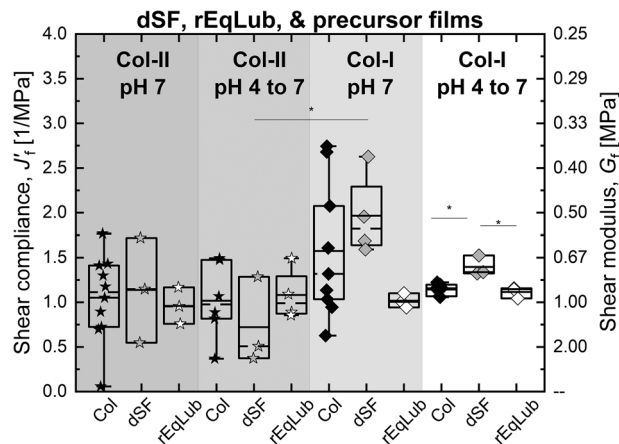


FIGURE 6 | Shear-film compliance of post-wash conditions for Col-II and Col-I films with adsorbed dSF or rEqLub. Box-and-whisker plot displays the mean (solid line), median (broken line), lower and upper quartiles (box) and minimum/maximum values excluding outliers with a coefficient of 1.5 (whiskers). Each data point is an independent replicate.

Considering that there was no significant adsorption of dSF on Col-I films, intriguingly, the “little to no” adsorption of dSF still affected the mechanical properties of the film. Given that film compliance is influenced by the molecular network’s cross-linking, water content, and protein density, among other factors, results suggest that dSF components altered Col-I films formed at pH 4 (Col-I pH 4 to 7) (Figure 6), thereby affecting their mechanical integrity. This effect was not evidenced on Col-II films, where dSF components were likely able to form denser layers.

2.10 | Control Experiments

Control experiments previously reported elsewhere [54] by our group consisted of quantifying m_f of dSF on bare Au surfaces, Au surfaces with SAM, and Au surfaces with SAM and a BSA blocking step, to gain insights into the role of collagen films in adsorbing and retaining dSF. In all these control conditions, m_f of dSF was significantly higher than m_f of dSF on the collagen films reported in this work, suggesting that the dSF adsorption on collagen films was specific to collagens and not to the underlying bare gold or SAM surfaces. The adsorption of BSA alone on Au and collagen films is shown in Figure S6. Adsorption of R-LUB on bare gold and SAM with glutaraldehyde is shown in Figure S7.

2.11 | Limitations and Outlook

Collagens at the surface of articular cartilage are present in fibrillar form [71, 72]. This higher-order structure provides mechanical strength to cartilage. It also yields a rougher surface topography that may influence interactions with SF components, thus controlling the supramolecular assembly of SF-derived films. While not the focus of this study, differences in the tribological performance may result from the additional, higher-order organization of collagens interacting with SF. Additionally, as with any other tissue, it is the collective contribution of individual extracellular matrix components that imparts each tissue’s unique function.

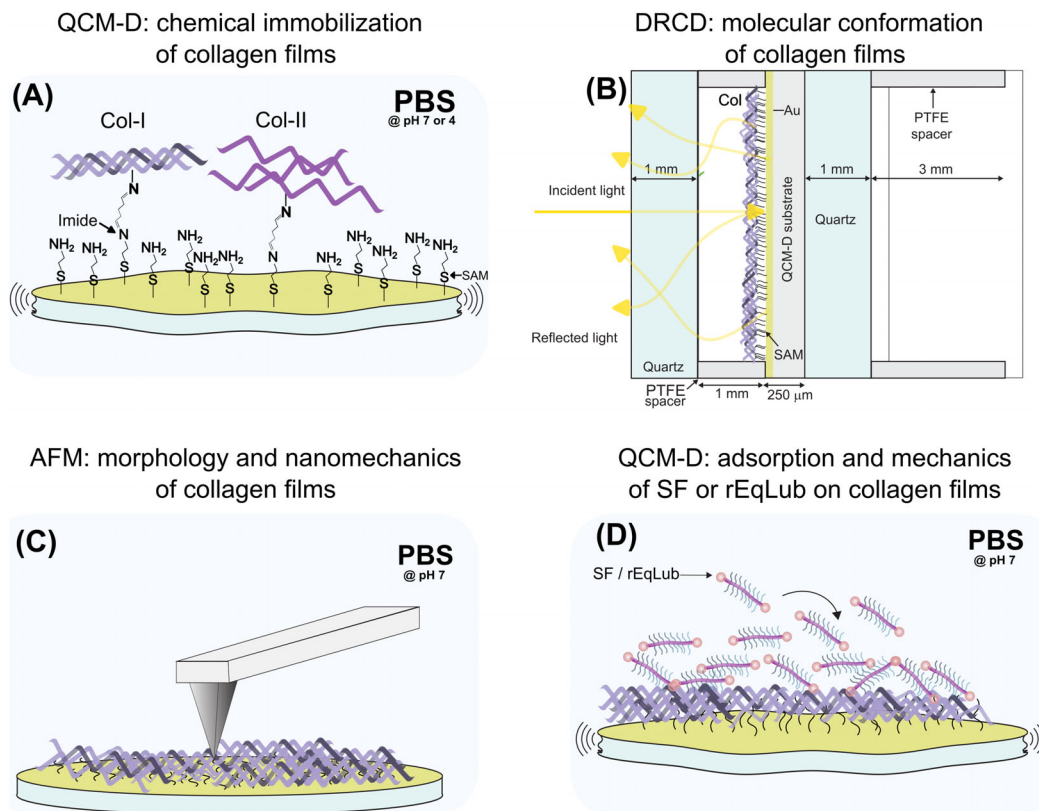


FIGURE 7 | Schematic summarizing experimental approaches and techniques, which include surface functionalization and characterization via QCM-D, DRCD and AFM. (A) Chemical immobilization of Col-II/Col-I to amine-terminated SAM surfaces through glutaraldehyde activation, in PBS at pH 7 or 4. (B) Assessing molecular conformation of collagen films via DRCD. (C) Collagen film morphology and mechanics investigated via AFM in liquid environment measurements of the collagen films. (D) Quantifying adsorption of SF/rEqLub to the collagen surfaces in PBS at pH 7 via QCM-D.

Here, films of isolated collagens were used to understand how collagen isoforms regulate SF and rEqLub adsorption and retention. However, at the articular cartilage surface, other key components are present [73]. Thus, incorporating mixtures of collagens and other cartilage extracellular matrix components, such as fibronectin or elastin [74, 75] are key for future studies.

Synergies are also present in SF components [13, 19, 67, 76]. Expanding on how the rEqLub used in this study interacts with other key SF molecular components will need to be addressed in future studies. However, since the sequence of the end domains of rEqLub are those of native equine lubricin [24], rEqLub interactions with model cartilage and native cartilage surfaces are not expected to differ from equine lubricin. The mucin domain consists of 59 synthetic KEPAPTP repeats and has been shown to lubricate in a dose-dependent manner [77].

3 | Conclusions

In this work, molecular films of Col-II and Col-I were chemically immobilized to self-assembled monolayers on gold, at two different initial pHs, 4 and 7. DRCD, QCM-D, and AFM showed differences in collagen conformations, nanomorphology, and mechanical properties among the films, with thicker, more compliant, and more structured films forming at neutral pH. Furthermore, collagen films were exposed to synovial fluid (dSF) and recombinant equine lubricin (rEqLub), to gain insight

into their ability to regulate the supramolecular assembly and retention of dSF and understand how isolated rEqLub interacts with Col-II and Col-I. Our results show that, when exposed to dSF, differences in adsorption of up to 100% between Col-II and Col-I were detected. This was not the case for rEqLub, which adsorbed, forming a discrete, non-continuous film (patches) with similar mass densities across all tested conditions, suggesting that lubricin alone does not drive full SF film assembly in our cartilage models. Our findings may offer insight into mechanisms underlying reduced mechanical performance in injured cartilage, potentially associated with Col-II loss and changes in the Col-II:Col-I ratio. Thus, the collagen isoform is a parameter that should not be overlooked when designing functional biological biomaterials intended for the AC-ECM (synthetic cartilage implants). Future efforts will focus on elucidating the tribological performance of these bio-interfaces.

4 | Experimental Section

4.1 | Materials

Phosphate buffer saline (PBS) (Gibco), ethanol (ACROS, absolute, 200 Proof, $\geq 99.5\%$), sodium dodecyl sulfate (SDS) (MP Biomedicals, ultrapure, $\geq 99\%$) were used as received. Base-piranha solution was prepared by mixing ammonium hydroxide (Chem-savers, 28%–30% pure), and hydrogen peroxide (PERDROGEN by Honeywell, MDL # MFCD00011333, $\geq 30\%$ w/w stabilized),

and water as follows: 6:1:1 by volume of water: ammonium hydroxide: hydrogen peroxide. Gold-coated silica quartz crystals were purchased from Quartz PRO with a resonance frequency of 5 MHz and a diameter of 14 mm. For gold surface functionalization, cysteamine hydrochloride (Sigma, $\geq 98\%$ titration) and glutaraldehyde (Sigma, 25% aqueous solution, Grade I) were used. Ultrapure water was collected from a Thermo-Fisher UV water purification system, with a resistivity of 18.2 M Ω •cm.

4.2 | Col-II and Col-I

Bovine collagen type II (Col-II, Sigma-Aldrich, Southern Biotech), bovine collagen type I (Col-I, Gibco) were used for collagen film formation. Col-II and Col-I solutions were stored as purchased at 4°C. The working concentration of 50 $\mu\text{g/mL}$ was prepared with PBS with a pH of 7.4 (referred to as PBS 7 or PBS at pH 7), or 4 (referred to as PBS 4 or PBS at pH 4). PBS with a pH of 4 was prepared by adding tiny acetic acid (0.1 M) droplets and was used within a month.

4.3 | Bovine Synovial Fluid

Bovine SF was stored at -80°C until use. To prepare working aliquots, frozen SF was thawed and centrifuged for 5 min at 6000 rpm to remove cell debris and large tissue aggregates, followed by a 25% v/v dilution in PBS as a thinning step to prevent QCM-D lines from clogging. Aliquots were stored at -80°C until the day of the experiment. This is referred to as dilute SF (dSF) in the text. dSF was always diluted and used with PBS at pH 7.

4.4 | Recombinant Equine Lubricin

A recombinantly-produced equine lubricin (rEqLub) was produced using codon scrambling, resulting in a maximum stock concentration of 20 mg/ml, with end-termini identical to those found in equine lubricin. The central mucin domain differs in sequence [24, 77]. The stock was diluted to 5 mg/ml in PBS and friction tested using a custom pin-on-plate tribometer [78] to confirm lubricity (details are provided in the SI, with results shown in Figure S8), as at this concentration, rEqLub has been shown to reach maximum lubricity in dose-response tribological tests [77]. For QCM-D testing, solutions were stored at -20°C and further diluted to a working concentration of 100 $\mu\text{g/mL}$ on the day of the experiment. rEqLub was always dissolved to working concentrations and used with PBS at pH 7.

All experiments using collagen, SF, and rEqLub were conducted in accordance with the procedures and guidelines of the Institutional Biosafety Committee at the University of California, Merced.

4.5 | Functionalization of Substrates with Self-Assembled Monolayers for QCM-D

Clean gold-coated crystals with a 5 MHz fundamental resonance frequency were rinsed with 2 wt% SDS, ultrapure water, and ethanol, respectively, and the process was repeated three times.

After rinsing, surfaces were dried with a gentle stream of N_2 , followed by an oxygen plasma cleaning step (Harrick Plasma, PDC-32G-115 V) for 2 min and stored in Petri dishes for immediate use.

Clean crystals were incubated in 0.088 M ethanolic solution of cysteamine hydrochloride at 4°C overnight. This process creates amine-terminated thiol-self-assembled monolayers on the gold surfaces and helps retain collagen, preventing collagen desorption due to Vroman effects [64] when exposing the collagen films to synovial fluid. The next day, surfaces were gently rinsed with ethanol to remove excess unbound NH_2 -terminated molecules and then dried under N_2 gas. Then, surfaces were incubated in 25% aqueous glutaraldehyde solution at room temperature for 30 min. This allowed imine formation between the aldehyde group of glutaraldehyde and the amine group of cysteamine hydrochloride (immobilization schematic shown in Figure 2A). After incubation, substrates were rinsed with water and then dried with an N_2 stream again. Functionalized surfaces were immediately loaded into the QCM-D fluidic cell.

After each use, crystals were cleaned in base piranha by submersion for ~ 30 s at 60°C . Next, crystals were rinsed with 2% SDS, water, and ethanol multiple times, then dried with a gentle stream of dry N_2 , finishing with a short plasma cleaning step. Each crystal was reused on average 5 times without loss of sensitivity. Crystals were discarded when the fundamental resonance frequency was below 4.95 MHz or the baseline drift tolerance was not met (< 2 Hz/hr).

4.6 | Quartz Crystal Microbalance with Dissipation (QCM-D)

QCM-D measurements were performed using a QCM-I mini (Micro Vacuum, Hungary) and a Q-Sense microbalance (Biolin, Sweden), using AT-cut quartz crystals with 5 MHz fundamental frequency coated with 50 nm of gold. All measurements were performed at 25°C controlled by an incorporated Peltier unit. The outlet channels of the QCM-I mini and Q-Sense were connected to peristaltic pumps (BQ80S Microflow Variable-Speed Peristaltic Pump and ISMATEC, respectively), which maintained a constant flow rate. All the QCM-D data presented in the main text come after reaching steady state.

The QCM-D experimental sequence began with a PBS (at pH 7 or 4) baseline (i.e., until a frequency dispersion of 2 Hz/hr or less over 2 min was achieved). Next, collagen solutions at concentrations ranging from 10 to 100 $\mu\text{g/mL}$ were flowed by applying suction with a peristaltic pump until the fluidic chamber was saturated. After saturation, the peristaltic pump was stopped, and changes in frequency signal and dissipation were collected until a steady state was achieved. This step is named “collagen pre-wash” in the text. PBS 7 or PBS 4 was used to wash loosely bound Col-II and Col-I molecules, and the signal was collected until a steady state was reached. This step is denoted as “Col-II/Col-I post-wash”. For experiments in which the collagen films were deposited at pH 4, a second PBS wash was performed at pH 7. In the text, data from this step is denoted as “pH 4 to 7”. Up to this point, the experimental sequence was the same for both types of experiments, collagen films exposed to dSF and rEqLub. Next, for

the rEqLub experiments, 100 µg/ml of rEqLub in PBS was flowed until a steady state was achieved (“rEqLub pre-wash”), followed by a PBS wash (“rEqLub post-wash”). On the other hand, for the dSF experiments, a blocking step with 10 mg/ml BSA in PBS at pH 7 was performed (“BSA pre-wash”), followed by a PBS wash at pH 7 (“BSA post-wash”). After this step, dSF was flowed, and data was collected until a steady state was achieved (“dSF pre-wash”). The experiments concluded with a wash with PBS at pH 7 (“dSF post-wash”). No BSA blocking step was used in the rEqLub conditions to facilitate quantitative comparison with literature [2, 11, 30, 69, 79–81].

The last 2 min of the steady-state values (Δf and ΔD) of Col-II or Col-I pre- and post-wash, dSF pre- and post-wash, and rEqLub pre- and post-wash were used to calculate the average mass densities, m_f , and thin film elastic compliances, J'_f , as described below. To calculate m_f of rEqLub adsorption on collagen films, frequency changes between the rEqLub pre- and post-washes steps were compared against the last PBS wash performed on the respective collagen film. For the case of dSF adsorption, the comparison was between dSF pre- and post-washes and BSA post-wash step. This approach allowed us to determine the areal mass of adsorbed dSF and retained dSF after wash, without accounting for the precursor films on functionalized gold surfaces. For shear film compliance, because deconvoluting the contribution of each layer is not straightforward, each condition was compared with the baseline, i.e., a combined system, for cases with rEqLub and dSF components adsorbed to the collagen films.

Each reported condition consists of at least three independent measurements, reported as the average $\pm$ standard error of the mean in Supporting Information tables, unless otherwise stated.

Control experiments with only Col-II or Col-I adsorption on bare gold (without any functionalization), rEqLub adsorption experiments on bare gold and on SAM-functionalized gold were also performed. dSF adsorption controls were omitted from this report, as these were previously published by our group elsewhere [54].

4.7 | QCM-D Data Analysis and Statistics

QCM-D data were analyzed using a combination of OriginLab Pro and Microsoft Excel. In this report, QCM-D data is shown as box-and-whisker plots, displaying the mean (solid line), median (broken line), lower and upper quartiles (box), and minimum/maximum values excluding outliers with a coefficient of 1.5 (whiskers). Each presented data point was used to calculate the mean and SEM. A data point is an independent measurement corresponding to the average of 2 min of data collected on the steady state of each experimental step, using pyQCM-BraTaDio [82].

To determine the impact of collagen type and formation pH, a two-way ANOVA of the post-wash conditions was performed using OriginLab. Next, post-hoc pairwise comparisons were performed using pairwise *t*-tests with Holm-Bonferroni-adjusted *p*-values to control the family-wise error rate, also using OriginLab. *, **, and *** indicate different levels of statistically significant differences, all detailed in the accompanying Supporting

Information “QCM-D_Pairwise_Comparisons.xlsx”. Group comparisons that are not labeled in the QCM-D related figures (Figures 2B,C, 5A,B, and 6) did not present statistically significant differences, or were not considered relevant comparisons for this study. However, comparisons between all groups of each figure are provided in Supporting Information “QCM-D_Pairwise_Comparisons.xlsx”.

4.8 | Modeling of QCM-D Data

Mass density (m_f) and shear elastic compliance (J'_f) of precursor (collagen) films and of adlayers (dSF/rEqLub) were calculated with the software pyQCM-BRATADIO [82]. m_f , using the Sauerbrey model, can be estimated as:

$$m_{f=} = c \cdot \frac{\Delta f_n}{n} \quad (1)$$

where c is the crystal mass sensitivity, typically $17.7 \text{ ng} \cdot \text{cm}^{-2} \cdot \text{Hz}^{-1}$ for a 5 MHz crystal, and Δf_n is the change in frequency for the overtone n . A linear regression was used to obtain the slope of the change in frequency versus all the available overtones, typically from the third to the 13th, as described in [54]. Shear modulus (G_f) is also reported as the inverse of shear compliance on a second y-axis. The Sauerbrey thickness, d_f , was estimated as follows: an effective dry mass density for the collagen films was calculated from [83]:

$$\rho_{eff} = \frac{m_{wet}}{\frac{m_{dry}}{\rho_{collagen}} + \frac{m_{H_2O}}{\rho_{H_2O}}} = \frac{2700 \frac{\text{ng}}{\text{cm}^2}}{\frac{490 \frac{\text{ng}}{\text{cm}^2}}{1.35 \frac{\text{g}}{\text{cm}^3}} + \frac{2210 \frac{\text{ng}}{\text{cm}^2}}{1 \frac{\text{g}}{\text{cm}^3}}} \approx 1.05 \frac{\text{g}}{\text{cm}^3}, \quad (2)$$

using the value of dry mass density for Col-II in PBS at pH 7 on gold, at 50 µg/ml, reported by Yuan et al. [30] (m_{dry}), as 490 ng/cm^2 , and of m_{wet} of 2700 ng/cm^2 , as obtained via QCM-D and reported for Col-II on bare Au (results in Supporting Information). A similar calculation for the case of Col-I was performed, using the Col-I value of 340 ng/cm^2 on Au at 48 µg/ml reported by Yuan et al. [37] obtaining values even closer to 1 g/cm^3 , i.e., to a film density dominated by water. Thus, the value obtained from Equation (2) was chosen to be used as ρ_{eff} across all films, assuming that it can be true for hydrated protein films with a 10% error or less. From this, a Sauerbrey thickness d_f in nanometers can be estimated as:

$$d_f = \frac{m_f}{\rho_{eff}} \quad (3)$$

The shear elastic compliance, J'_f was calculated using:

$$\frac{\Delta \Gamma}{-\Delta f} \approx J'_f 2\pi n f_0 \eta_{bulk}, \quad (4)$$

as derived by Du and Johannsmann [84], where f_0 is the fundamental resonance frequency, η_{bulk} is the viscosity of the media, and $\Delta \Gamma$ is the change in half-band half-width. It is related to changes in dissipation D as follows:

$$\Gamma = \frac{D f_n}{2}, \quad (5)$$

with $f_n = n \cdot f_0$ as the resonance frequency of the n^{th} overtone. In this approximation, it is assumed that the density of the bulk fluid (PBS) and the immobilized/adsorbed films are similar, which, based on Equation (2), is likely to be true within 10% error. Representative QCM-D traces for changes in frequency and dissipation of each condition are shown in Supporting Information (Figure S9 (dSF experiments) and Figure S10 (rEqLub experiments)).

4.9 | Immobilization of Collagens for DRCD and AFM

Immobilization of Col-I and Col-II for DRCD and AFM analysis was performed outside the QCM-D fluidic chamber as follows: a clean QCM-D crystal was functionalized with a SAM and activated with glutaraldehyde, as for QCM-D measurements. After the glutaraldehyde activation step, the crystals were rinsed with ultrapure water, followed by complete immersion in a clean glass jar containing 50 $\mu\text{g/mL}$ protein solution at the corresponding pH for 2 h at 4°C, followed by a PBS wash at their respective pH, and ending with a PBS 7 wash for the case of pH 4 conditions.

4.10 | DRCD

DRCD spectra were collected on a circular dichroism spectrometer J-1500 (JASCO, Japan) operating in diffuse reflectance mode under a nitrogen atmosphere. Samples were placed in a sample holder and immersed in PBS (pH 7). A total of 10 spectra accumulations were collected and averaged, with a digital integration time of 4 s, a bandwidth of 4 nm, and a scanning speed of 100 nm/min. Spectra were baseline-corrected, smoothed with a Savitzky-Golay filter (polynomial order 1, frame length 9) [85], and zero-endpoint corrected at the long-wavelength limit of each window. Data is presented as mean residue ellipticity (*MRE*), with units of $\text{mdeg} \cdot \text{m}^2 \cdot \text{mol}^{-1} \cdot \text{residue}^{-1}$, calculated from:

$$MRE = \frac{\theta \cdot M_w \cdot 10^6}{2 \cdot m_f \cdot N_r}, \quad (6)$$

where θ is the measured ellipticity in millidegrees, m_f is the surface-specific mean Sauerbrey mass in ng/cm^2 , M_w is the protein molecular weight in g/mol, and N_r is the total residue count per molecule. These values are shown in Table S9.

For collagen samples with the spectra exhibiting the canonical triple-helix band structure, the ratio between the magnitudes of the positive peak at 222 nm and the negative peak at 197 nm [86] is reported as:

$$R_{\text{pn}} = \frac{\theta_{p,222}}{\theta_{n,197}}. \quad (7)$$

Because the R_{pn} is a ratio of two ellipticities at two wavelengths, it is mathematically invariant under any scalar multiplier of the spectrum (effective concentration, path length, film thickness) and is therefore in principle well-defined for surface-confined DRCD films. It should be noted that R_{pn} from DRCD is not numerically comparable to published transmission-CD R_{pn} values because DRCD and transmission CD use different

baseline-subtraction conventions (bare-substrate / zero-endpoint vs solvent blank), and the additive residual baseline does not cancel in a ratio. R_{pn} is therefore reported for relative comparison within the present dataset only. For conditions where the canonical triple-helix was not present, the comparisons were limited to spectral shape and *MRE* magnitudes at the diagnostic wavelengths.

4.11 | CD

The bulk collagen samples were measured in PBS solution at a protein concentration of 0.01 mg/mL, measured in transmission using quartz cuvettes with the pathlength of 1 mm. The samples were measured as the average of three spectra accumulations from 250–190 nm at 25°C, and were subsequently baseline and zero-endpoint corrected, and converted to *MRE*. R_{pn} was also calculated according to Equation (7).

4.12 | AFM

A Nanowizard Pure AFM (Bruker, USA) was used for imaging immobilized protein films immersed in PBS at pH 7. ScanAsyst-Fluid probes (Bruker, USA), with a nominal normal spring constant of 0.7 N/m were used. Cantilevers were V-shaped, gold-coated on the backside. QI mode was used to obtain height maps of collagen films, as it relies on a fixed tip-sample force setpoint, allowing imaging of soft surfaces with minimal damage. For the images of collagen films with rEqLub, contact mode imaging was performed, to visualize possible differences in the tip-sample interactions when scanning on rEqLub molecules. Setpoint was 0.4 nN, with a tip velocity of 11.3 $\mu\text{m/s}$ and a line rate of 1 Hz for 512 lines. All topographical images were processed using Gwyddion [87] version 2.6.7. Second-degree polynomial row alignment and horizontal scar corrections were applied to the raw images. Mean-squared roughness (RMS) of height irregularities is reported as calculated using Gwyddion.

Using the same type of probe, force-volume measurements were performed to further extract mechanical maps from at least two $2 \times 2 \mu\text{m}^2$ areas per condition, consisting of 16 384 force-distance curves per $2 \times 2 \mu\text{m}^2$ area. In-run of force-distance profiles were fitted with the Hertz [88, 89] model to estimate the Young's modulus, with JPKSPM Data Processing version 8.1.62.

Particularly, for every force-distance curve, the entire in-run of the force measurements was considered, with the 4%–20% region of the full run analyzed, where 0% corresponds to the last point of the in-run and 100% corresponds to the starting point, preventing measuring artifacts due to the effect of the gold substrate in the mechanical characterization of the films. One of the flaws of using this method, particularly with “sharp” AFM tips, is the unknown area of contact, added to the uncertainty in the value of Poisson's ratio, crucial in the Hertz model, which was assumed as 0.5, which for soft materials is not necessarily the case [90, 91], even though it can be a first approximation for highly hydrated materials. To simplify direct comparison of QCM-D shear-compliance and the AFM extracted normal Young's modulus (*E*), normal compliance *S* values are reported as the inverse of the Young's modulus as well, i.e., $S = 1/E$. *E*

distributions for all conditions are reported as histograms with a bin size of 0.33 MPa, overlapping measurements on the probed areas. Mean and standard deviation (STD) for each distribution were calculated using a Gaussian fit in OriginLab. For each condition, the average of the means is reported as E_{avg} , along with the average of their STDs. Described characterization techniques are schematized (Figure 7).

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Conflicts of Interest

M.J.P. and H.L.R. are named inventors on patents assigned to Cornell University and related to the production of recombinant lubricins and mucins. The remaining authors, D.R.J.P., N.L.M., S.T.A., Y.W., L.V., S.M.W., K.A.C., U.H., L.J.B., K.D., R.C.A.E. declare no competing interests.

Data Availability Statement

The data that support the findings of this study are available from the corresponding author upon reasonable request.

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Supporting Information

Additional supporting information can be found online in the Supporting Information section.

Supporting File 1: adfm77839-sup-0001-SuppMat.pdf.

Supporting File 2: adfm77839-sup-0002-SuppMat.xlsx.